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## **CeMM study reveals how a master regulator of gene transcription operates**

**The human Mediator complex is selectively required for cell-type-specifying transcription**

**Using targeted protein degradation technology, researchers at CeMM, the Research Center for Molecular Medicine of the Austrian Academy of Sciences, set out to understand the primary role of a key regulator of transcription, the human Mediator complex. Mediator, a 30 subunit molecular machine, was thus far positioned as a factor that is generally required for the transcription of all protein-coding genes in human cells. In contrast, the recent study by Georg Winter's group at CeMM discovered that Mediator particularly safeguards the transcription of a small set of cell-type-specifying genes. In collaboration with Patrick Cramer's lab at the Max Planck Institute for Biophysical Chemistry in Göttingen, the scientists revealed mechanistic details of how Mediator enables cells to robustly exert their dedicated functions. The study "Selective Mediator dependence of cell-type-specifying transcription" was published in *Nature Genetics*, on 1 June 2020.**

(Vienna, 1 June 2020) Georg Winter's lab at CeMM uses chemical tools to understand the molecular basis of gene control and aberrations thereof in cancer. To do so, the team combines particularly fast-acting protein ablation systems ("degradation Tag"/ "dTAG", co-developed by Georg Winter in Jay Bradner's lab at the Dana Farber Cancer Institute, in Boston) with precise and unbiased measurements of gene activity at high kinetic resolution. Together with Patrick Cramer's lab and others, the international team of scientists now unveiled how the Mediator molecular machine mechanistically directs gene activation, which had long remained poorly understood owing to challenges in experimental manipulation.

Mediator was identified over 30 years ago as a molecular bridge that allows DNA-binding proteins, called transcription factors, to communicate with the cellular gene-copying machine, called RNA polymerase II (Pol II), to activate target genes. Understanding the direct role of Mediator in this process requires technologies that rapidly block Mediator function and measure changes in Pol II activity within the subsequent minutes. Using targeted protein degradation, scientists in the Winter lab now managed to rapidly remove individual parts of the Mediator complex to ask the question, whether the copying of all human genes depends to the same extent on Mediator integrity. In contrast to the existing paradigm, the study first-authored by CeMM PhD student Martin Jäger suggests that Mediator is not generally required for transcription of all genes. Rather, the data imply that Mediator selectively safeguards the expression of a small set of genes, which form densely connected regulatory circuits to instruct cell-type-specific functions.

By visualizing Pol II enzymes in cells, the scientists observed that Mediator nucleates large clusters of transcription machinery components that are thought to form around regulatory DNA regions, called super-enhancers. These super-enhancers direct the expression of cell-type-specific genes by physically touching their target genes, which are often millions of DNA bases away. When Mediator was degraded, the large Pol II clusters rapidly disappeared, but super-enhancers still seemed to touch their target genes, indicating that Mediator was not required to maintain these DNA contacts.

Together with experts from the Cramer lab in Göttingen, however, the team observed dramatic changes in the dynamics of Pol II turnover at super-enhancer-driven genes after Mediator loss. The data suggested

that Mediator clusters drove highly efficient recruitment of Pol II enzymes to these cell-type-specific genes, allowing their copying at remarkable efficiency. At the same time, the majority of other genes seemed to be surprisingly mildly affected by acute Mediator loss, which made the scientists suspect that they undergo some unexpected type of compensatory boost.

The team went on to address this compensatory mechanism, and found that promoter-proximal pausing of Pol II was globally reduced in response to Mediator loss. Pausing is a phenomenon, where Pol II enzymes are held back for several minutes right after they started copying a gene. The waiting Pol II constitutes a roadblock that prevents the passing of other Pol II enzymes behind it, thus limiting the number of RNA copies produced in a given timespan. After Mediator degradation, the scientists observed that the main factor responsible for signaling pause release, called cyclin dependent kinase 9 (CDK9), more efficiently bound to DNA in cells and more actively tagged its protein targets with a phosphate group. Chemically blocking CDK9 activity made the transcriptional defects of Mediator ablation less selective for super-enhancer target genes, which highlights that the uncovered CDK9 activation shapes how cells react to acute Mediator loss. This unexpected finding suggested that pausing may have evolved as a buffering capacity to rapidly react to and partially compensate acute defects in Pol II recruitment.

The study opens up new avenues to understand the composition of Mediator-nucleated Pol II clusters and how CDK9 is so efficiently activated in response to transcriptional stresses. Furthermore, Georg Winter imagines future possibilities to perhaps tackle diseases, where Mediator function went awry: *“Very preliminarily, our work might also have revealed some potential for Mediator as a drug target and next steps might be to develop direct degrader molecules against this complex.”*

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#### **Attached pictures:**

1. Georg Winter and Martin Jäger (© Laura Alvarez / CeMM)
2. Illustration: Visualizing gene expression dynamics revealed how Mediator directs the delivery of the cellular gene-copying machine (RNA polymerase II) particularly to a subset of cell identity genes (© Martin Jäger / CeMM)

**The study** “Selective Mediator dependence of cell-type-specifying transcription” was published in *Nature Genetics* on 1 June 2020. DOI: 10.1038/s41588-020-0635-0.

#### **Authors:**

Martin G. Jaeger, Björn Schwalb, Sebastian D. Mackowiak, Taras Velychko, Alexander Hanzl, Hana Imrichova, Matthias Brand, Benedikt Agerer, Someth Chorn, Behnam Nabet, Fleur M. Ferguson, André C. Müller, Andreas Bergthaler, Nathanael S. Gray, James E. Bradner, Christoph Bock, Denes Hnisz, Patrick Cramer,\* Georg E. Winter\*

\*co-senior authors

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**Georg Winter** obtained his PhD from the Medical University of Vienna, working on elucidating the mechanism of action of anti-neoplastic drugs under the supervision of Prof. Giulio Superti-Furga at CeMM. He specialized in proteomics as well as chemical genetic approaches to identifying drug resistance mechanisms and on mechanistically elucidating synergistic drug combinations. He continued his training in chemical biology, working as a postdoctoral fellow with Dr. James Bradner at the Dana Farber Cancer Institute/Harvard Medical School. There, he innovated a generalizable pharmacological solution to in vivo target protein degradation and applied this strategy to the study of leukemic gene regulation. Georg Winter was recruited as a CeMM Principal Investigator in June 2016. His lab develops and applies methods for target protein degradation with the ultimate goal of understanding and disrupting oncogenic transcriptional circuits. To that end, the Winter laboratory combines phenotypic drug screens, chemical genetics and drug-target identification approaches with holistic measurements of global gene activity and genome structure. The ultimate goal of the research conducted in the Winter laboratory is to connect basic research in gene regulation and the ubiquitin-proteasome system with functional genomics and chemical probe development to develop novel and personalized therapeutic paradigms.

The mission of **CeMM Research Center for Molecular Medicine of the Austrian Academy of Sciences** is to achieve maximum scientific innovation in molecular medicine to improve healthcare. At CeMM, an international and creative team of scientists and medical doctors pursues free-minded basic life science research in a large and vibrant hospital environment of outstanding medical tradition and practice. CeMM's research is based on post-genomic technologies and focuses on societally important diseases, such as immune disorders and infections, cancer and metabolic disorders. CeMM operates in a unique mode of super-cooperation, connecting biology with medicine, experiments with computation, discovery with translation, and science with society and the arts. The goal of CeMM is to pioneer the science that nurtures the precise, personalized, predictive and preventive medicine of the future. CeMM trains a modern blend of biomedical scientists and is located at the campus of the General Hospital and the Medical University of Vienna. [www.cemm.oeaw.ac.at](http://www.cemm.oeaw.ac.at)

For **further information** please contact

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**Laura Alvarez**

Social Media and Communications Manager

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**CeMM**

Research Center for Molecular Medicine  
of the Austrian Academy of Sciences

Lazarettgasse 14, AKH BT 25.3

1090 Vienna, Austria

Phone +43-1/40160-70 057

Fax +43-1/40160-970 000

[lalvarez@cemm.oeaw.ac.at](mailto:lalvarez@cemm.oeaw.ac.at)

[www.cemm.at](http://www.cemm.at)